Amendments to the Specification:

Please replace the paragraph beginning at page 1, line 5, with the following rewritten paragraph:

This application is a continuation-in-part of application S.N. 10/603,000, filed June 23, 2003, which is a continuation-in-part of application S.N. 09/727,361, filed November 29, 2000, now U.S. Patent 6,657,048, issued December 2, 2003, which is a continuation-in-part of application S.N. 09/415,278, filed October 8, 1999, now U.S. Patent 6,180,357 B1, the contents of each of which are herein incorporated by reference.

Please replace the paragraph beginning at page 32, line 7, with the following rewritten paragraph:

In order to obtain better resolution of the binding entities, and to further characterize the proteins, 2-dimensional electrophoresis was carried out. Total membrane proteins (75-200 µg) prepared as described above, were precipitated using the PlusOne 2-D Clean Up kit (Amersham; Baie D'Urfé, QC), and then resuspended in rehydration buffer containing ampholytes in the pH range 3-10. Samples were centrifuged to remove particulate material, and then loaded onto IPG strips (Amersham; Baie D'Urfé, QC) in the presence of a rehydration solution. Proteins were focussed using the following protocol: 16 hr for rehydration; 500 V, 250 Vhrs, 1000 V, 500 Vhrs; 5000 V, 7500 V hrs. The strip was then removed from the strip holders, and incubated in an SDS-PAGE equilibration buffer. After 15 min, the strip was placed on the top of an 8 percent gel, and sealed with an agarose solution. Prestained MW markers were loaded beside the strip. Electrophoresis was

carried out at 100 V for 10 min, followed by 65 min at 150 V. One of the gels was fixed for 30 min with 10 percent methanol/7 percent acetic acid, and then stained with the fluorescent dye SYPRO [[Ruby^{TM}]] Ruby protein gel stain (Molecular Probes, Eugene, OR). Protein spots were visualized under UV light. From a second and third gel, proteins were transferred from the gels to PVDF (Millipore) membranes by electroblotting for 16 hr at 40 V. Quantitative transfer was assessed by determining the complete transfer of the prestained markers from the gel to the membrane.

Please replace the paragraph beginning at page 33, line 11, with the following rewritten paragraph:

Figure 3a demonstrates the Western blot obtained from MB-468 membranes incubated with H460-16-2. Two distinct binding spots are observed, with molecular weights corresponding with those obtained by 1-dimensional electrophoresis. One is observed at a MW of approximately 80-90 kD according to the MW standards, and is in the acidic portion of the gel with an estimated pI of 3 – 4. The second spot is in the MW range of 120-150 kD according to the MW standards, and has a pI more basic than the 80-90 kD protein. Figure 3b demonstrates the Western blot obtained from membranes incubated with the isotype control antibody. No spots were visible on this blot, indicating that the binding of H460-16-2 was not due to non-specific binding. Figure 3c shows a SYPRO [[Ruby^{TM}]] Ruby protein gel stain stained 2D-gel of MB-468 membrane proteins. Note that when a similar Western blot is run with MB-231 membranes, only the 80-90 kD spot is observed (Figure 4).

Please replace the paragraph beginning at page 35, line 17, with the following rewritten paragraph:

Three mg of total membrane protein from MB-231 cells was incubated with H460-16-2 chemically crosslinked beads in 0.1 M sodium phosphate buffer, pH 7.4 containing 0.1 percent Tween-20, 5 percent glucose, 5 percent mannose, 5 percent galactose and protease inhibitors at 4°C for 4 hr. After incubation, the immunoprecipate was washed 3 times in PBS containing 150 mM NaCl and 0.1 percent Tween-20. Protein was eluted from the beads by incubating the H460-16-2-crosslinked beads with 0.1 M citrate, pH 3 for 4 min. Eluted protein was stored at -80°C. Immunoprecipitated protein from 3 mg of protein was loaded onto a single lane of an 8 percent non-reducing SDS-PAGE gel. A sample of pre-stained molecular weight markers (Biorad; Mississauga, ON) was run in a reference lane. The sample was separated by electrophoresis at 100 V for 10 min, followed by 65 min at 150 V. Proteins were stained with SYPRO [[Ruby™]] Ruby protein gel stain.